



## International Journal of Surgery

Volume 24, Part B, December 2015, Pages 143-146

Review

# Biological mechanism of shockwave in bone

Jai-Hong Cheng <sup>a, b</sup>, Ching-Jen Wang <sup>a, c</sup>  

 **Show more**

<https://doi.org/10.1016/j.ijisu.2015.06.059>

[Get rights and content](#)

Under an Elsevier [user license](#)

[open archive](#)

### Highlights

- ESWT is a novel method in musculoskeletal disorders and other disease.
- The biologied effects of ESWT may be through mechanotransduction.
- Applications of ESWT are increasing.
- ESWT promotes tissue regeneration, wound healing, angiogenesis, bone remodeling, and anti-inflammation.

### Abstract

Shockwave is a rapid, short duration acoustic wave that carries energy and can propagate through tissue medium. This kind of physical force can be a mechanical stimulus that

induces biological effects in living tissue. [Extracorporeal shockwave therapy](#) (ESWT) acts as a mechanical stimulus which promotes biological healing processes through a [mechanotransduction](#). The biological effects of ESWT are reported such as [tissue regeneration](#), wound healing, [angiogenesis](#), [bone remodeling](#), and anti-inflammation. Until now, however, little is known about the basic mechanism of action of this type of therapy. This article describes the molecular mechanism on the current status of ESWT with pre-clinical and clinical applications for treating disorders in bone.

[< Previous](#)

[Next >](#)

## Keywords

Biological mechanism; Shockwave; Bone

## 1. Introduction

[Extracorporeal shockwave therapy](#) (ESWT) has been used for [musculoskeletal disorders](#) over 30 years. Several studies had investigated the effects of shockwave therapy on [fracture healing](#) and [articular cartilage](#) in animal and human experiments [1], [2], [3], [4]. The positive effect of shockwave in promoting bone healing was demonstrated in both acute fracture and chronic [non-union](#) in animal experiments [2], [5], [6]. Despite of clinical success, the working mechanism of ESWT in bone healing has not been fully established. The mechanism of ESWT is suggested through [mechanotransduction](#) to induce the reaction of the bone lacunae-canalicular network to tensile, shear and compression forces [7], [8]. It was speculated that shockwave produced [micro-fracture](#) that in turn causes hematoma formation and subsequent callus formation and eventual fracture healing [9], [10]. However, there were insufficient data to scientifically substantiate the theory. In fact, other studies demonstrated that ESWT significantly promotes bone healing after fracture and tendon to bone healing in bone tunnel [10], [11].

The use of ESWT has recently expanded from skeletal disorders to non-skeletal diseases such as acute and chronic wound healing, [diabetic foot ulcers](#), ischemic myocardiac disease and erectile [sexual dysfunction](#) [12], [13], [14], [15], [16]. The clinical results showed some differences among different series, but the great majority of the reported series showed positive effects of ESWT in different skeletal and non-skeletal disorders

up to 2014. Despite of clinical success of ESWT in different diseases, the working mechanism of ESWT has not been fully established.

## 2. ESWT treatment on tendon to bone

To explore the biological mechanism of ESWT in biological tissues, many studies attempted to elucidate the mechanism of ESWT from the basic science study and translate into clinical application [10], [13], [17], [18]. In an experiment in rabbits, Wang CJ and his colleagues reported that application of ESWT caused the ingrowth of **neovascularization** associated with **up-regulation** of angiogenic and osteogenic growth factors including **endothelial nitric oxide synthase** (eNOS), vessel **endothelial growth factor** (VEGF), **proliferating cell nuclear antigen** (PCNA), and bone morphogenic protein-2 (BMP-2) at the tendon–bone junction of the **Achilles tendon** in rabbits [3], [10]. The increase in neo-vessels began to rise in one week after ESWT application, and reached the plateau in four weeks and then, persisted through twelve weeks. The up-regulation of eNOS, VEGF and **BMP-2** showed significant increases in one week, and reached the peak values at 12 weeks, then slowly returned to baseline data at the end of 12 weeks. The change of PCNA starts to rise at 1 week after ESWT application, and the highest value was observed at 12 weeks. The results indicate that ESWT causes the ingrowth of neovascularization beginning in one week after treatment and such effect persisted beyond 12 weeks after treatment. This is also supported by the persistent elevation of PCNA at 12 weeks after treatment although the maximal changes on the effects of eNOS, VEGF and BMP-2 returned to baseline value at 12 weeks after treatment. This is the first time that the biological responses had been conclusively shown in the literature. Subsequently, several studies reported similar results with various target tissues in biological tissues including **non-union** of the long bone fractures [11], [19].

## 3. Treatment of osteonecrosis of the hip

Some studies examined the biological mechanism of ESWT in the treatment of **osteonecrosis** of the hip joint [20], [21], [22]. The patients with osteonecrosis of the **femoral head** were treated with ESWT [23]. In histopathological examination, ESWT treated hips showed significantly more viable bone and less necrotic bone, higher cell concentration and more cell activities including **phagocytosis** than those without ESWT before hip replacement. In molecular expression analysis, ESWT showed significant increases in vWF, VEGF, CD 31, Wnt3 and PCNA, and decreases in **VCAM** and Dickkopf-1 (DKK-1) than those without ESWT before surgery [23]. In another study, **bone marrow stromal cells** (BMSCs) were harvested from the bone marrow cavity of the **proximal**

femur in six patients with osteonecrosis [22]. There were significant increases in cell proliferation, VEGF, alkaline phosphatase, BMP-2, runt-related transcription factor 2 (RUNX2) and osteoclast in mRNA expressions in the shockwave group. These results demonstrated that ESWT significantly enhances the angiogenic and osteogenic effects of the BMSCs mediated through the nitric oxide pathway in hips with osteonecrosis, and these findings gave some insights into the biological effects of ESWT in bone [22].

Recently, additional studies reported the use of shockwave in osteonecrosis of the hip in 35 patients with 47 hips with special attention to nitric oxide (NO<sub>3</sub>) pathway. At 12 months, 83% showed improvement and 17% unimproved. At 1 month, ESWT-treated cases demonstrated significant elevations of angiogenic growth factors including NO<sub>3</sub>, VEGF, vWF and FGF basic and a decrease in TGF-β1. There were also significant increases in osteogenic factors including BMP-2, osteocalcin, alkaline phosphatase and IGF and a decrease in DKK-1 at one month after treatment. These changes in peripheral blood tests only lasted for 1 month post-shockwave [21].

#### 4. ESWT on osteoarthritis

The biological effects of ESWT in osteoarthritis (OA) of the knee has been studied, and the results showed that application of ESWT to the subchondral bone of the medial tibia condyle showed time dependent, site specific chondroprotective effects in the initiation of OA changes of the knee in rats [19], [24], [25]. There were significant increases of VEGF, BMP-2, and osteocalcin in the subchondral bone as compared to the control at week 2, 4, 8, and 12. The most beneficial effects of ESWT in the OA knee occurred at 4 weeks after shockwave application. Such effects seemed to continue until 12 week [19]. Recent research demonstrated osteoporosis (OP) increased the severity of cartilage damage in osteoarthritis of the knee. ESWT showed effectiveness in the reduction of osteoporotic osteoarthritis of the knee in rats. In immunohistochemical analysis, DKK-1 significantly increased, but VEGF, PCNA, and BMP-2 decreased in groups with osteoarthritis, osteoporosis, and osteoarthritis plus osteoporosis relative to the sham group, and ESWT significantly reversed the changes of osteoarthritis of the knee [26].

#### 5. ESWT treatment on bone to cartilage

Many studies reported intensive osteochondrogenesis in segmental femoral defects after shockwave treatment, but no shockwave-induced crack or micro-damage was noted on bone [27], [28], [29]. Therefore, shockwave-augmented bone formation may be attributed to shockwave-sensitive osteogenesis, rather than damage to the bone architecture. However, some reports showed high-energy ESWT *in vivo* affected the structural

integrity of [articular cartilage](#) [30]. Tenascin-C and Chi3L1 expressions showed signals indicating reorganization in [matrix protein](#) composition connected to [cartilage injury at 10 weeks](#) after high-energy ESWT [30]. This study speculated the possibility of long-term degenerative effects of ESWT on cartilage. Other studies demonstrated that [TGF-β1](#), [BMP-2](#) and VEGF regulated the [mechanical stimulation](#) of [fracture healing](#) [31], [32]. Recent studies showed that shockwave promotion of fracture healing coincided with increased [TGF-β1](#) and [BMP-2](#) expressions and [extracellular signal-regulated kinase](#) (ERK) and [P38 kinase](#) in callus [27], [28], [29]. A growing number of studies demonstrated that the increases of systemic osteogenic factors reflecting a local stimulation of bone formation during fracture healing [33], [34], [35]. Current studies reported the biological mechanism of ESWT in bone healing, and investigated that ESWT accelerates fracture healing with the improvement of [neovascularization](#) and enhancement of [angiogenesis](#) and osteogenesis growth factors including eNOS, VEGF, PCNA and [BMP-2](#) [10]. Other studies showed that ESWT triggers the cascade of angiogenic and osteogenic transcription factors (Cbfal/Runx2, HIF-1α and VEGF) in [osteoblast](#) cells [36], [37]. Meanwhile, evidence showed that shockwave energy induces [nitric oxide](#) (NO) elevation that promotes proliferation and differentiation of human osteoblasts [38].

## 6. Conclusion

ESWT is a non-invasive [therapeutic modality](#) with effectiveness, convenience, and safety. ESWT can replace surgery with no surgical risks in many orthopedic disorders including [non-union](#) of long bone fracture. The complication rates are low and negligible. However, the exact biological mechanism of [shockwave therapy](#) in [bone healing](#) is still unknown. Additional studies such as [proteomics](#), [transcriptome](#) and [next generation sequencing](#) technologies are needed to elucidate the biological mechanism of ESWT in biological bone tissues.

## Ethical approval

None.

## Funding

None.

## Author contribution

Ching-Jen Wang, conception and design, writing, final proof of the manuscript.

Jai-Hong Cheng, conception and design, writing, final proof of the manuscript.

## Conflict of interest

None.

## Guarantor

Ching-Jen Wang.





Jai-Hong Cheng.

[Special issue articles](#)

[Recommended articles](#)

[Citing articles \(20\)](#)

## References

- [1] M. Delius, K. Draenert, Y. Al Diek, Y. Draenert  
**Biological effects of shock waves: in vivo effect of high energy pulses on rabbit bone**  
Ultrasound Med. Biol., 21 (1995), pp. 1219-1225  
[Article](#)  [Download PDF](#) [View Record in Scopus](#) [Google Scholar](#)
- [2] G. Haupt, A. Haupt, A. Ekkernkamp, B. Gerety, M. Chvapil  
**Influence of shock waves on fracture healing**  
Urology, 39 (1992), pp. 529-532  
[Article](#)  [Download PDF](#) [View Record in Scopus](#) [Google Scholar](#)
- [3] C.J. Wang, F.S. Wang, K.D. Yang, L.H. Weng, C.C. Hsu, C.S. Huang, L.C. Yang  
**Shock wave therapy induces neovascularization at the tendon-bone junction. A study in rabbits**  
J.Orthop. Res.: Off. Publ. Orthop. Res. Soc., 21 (2003), pp. 984-989  
[Article](#)  [Download PDF](#) [View Record in Scopus](#) [Google Scholar](#)
- [4] E.J. Johannes, D.M. Kaulesar Sukul, E. Matura  
**High-energy shock waves for the treatment of nonunions: an experiment on dogs**  
J. Surg. Res., 57 (1994), pp. 246-252  
[Article](#)  [Download PDF](#) [View Record in Scopus](#) [Google Scholar](#)

- [5] R.W. Hsu, C.L. Tai, C.Y. Chen, W.H. Hsu, S. Hsueh  
**Enhancing mechanical strength during early fracture healing via shockwave treatment: an animal study**  
Clin. Biomech., 18 (2003), pp. S33-S39  
[Article](#)  [Download PDF](#) [View Record in Scopus](#) [Google Scholar](#)
- [6] C.J. Wang, H.Y. Huang, H.H. Chen, C.H. Pai, K.D. Yang  
**Effect of shock wave therapy on acute fractures of the tibia: a study in a dog model**  
Clin. Orthop. Relat. Res. (2001), pp. 112-118  
[CrossRef](#) [View Record in Scopus](#) [Google Scholar](#)
- [7] E.H. Burger, J. Klein-Nulend  
**Mechanotransduction in bone—role of the lacuno-canalicular network**  
FASEB J.: Off. Publ. Fed. Am. Soc. Exp. Biol., 13 (Suppl. 1) (1999), pp. S101-S112  
[CrossRef](#) [View Record in Scopus](#) [Google Scholar](#)
- [8] H. van der Worp, I. van den Akker-Scheek, H. van Schie, J. Zwerver  
**ESWT for tendinopathy: technology and clinical implications**  
Knee Surg. Sports Traumatol. Arthrosc.: Off. J. ESSKA, 21 (2013), pp. 1451-1458  
[CrossRef](#) [View Record in Scopus](#) [Google Scholar](#)
- [9] J.A. Ogden, A. Toth-Kischkat, R. Schultheiss  
**Principles of shock wave therapy**  
Clin. Orthop. Relat. Res. (2001), pp. 8-17  
[View Record in Scopus](#) [Google Scholar](#)
- [10] C.J. Wang, F.S. Wang, K.D. Yang  
**Biological effects of extracorporeal shockwave in bone healing: a study in rabbits**  
Arch. Orthop. Trauma Surg., 128 (2008), pp. 879-884  
[CrossRef](#) [View Record in Scopus](#) [Google Scholar](#)
- [11] C.J. Wang, K.D. Yang, J.Y. Ko, C.C. Huang, H.Y. Huang, F.S. Wang  
**The effects of shockwave on bone healing and systemic concentrations of nitric oxide (NO), TGF-beta1, VEGF and BMP-2 in long bone non-unions**  
Nitric Oxide: Biol. Chem. Off. J. Nitric Oxide Soc., 20 (2009), pp. 298-303  
[Article](#)  [Download PDF](#) [View Record in Scopus](#) [Google Scholar](#)
- [12] R. Mittermayr, V. Antonic, J. Hartinger, H. Kaufmann, H. Redl, L. Teot, A. Stojadinovic, W. Schaden



**Extracorporeal shock wave therapy (ESWT) for wound healing: technology, mechanisms, and clinical efficacy**

Wound Repair Regen.: Off. Publ. Wound Heal. Soc. Eur. Tissue Repair Soc., 20 (2012), pp. 456-465

[View Record in Scopus](#) [Google Scholar](#)

[13] C.J. Wang

**Extracorporeal shockwave therapy in musculoskeletal disorders**

J. Orthop. Surg. Res., 7 (2012), p. 11

[CrossRef](#) [View Record in Scopus](#) [Google Scholar](#)

[14] C.J. Wang, Y.R. Kuo, R.W. Wu, R.T. Liu, C.S. Hsu, F.S. Wang, K.D. Yang

**Extracorporeal shockwave treatment for chronic diabetic foot ulcers**

J. Surg. Res., 152 (2009), pp. 96-103

[Article](#)  [Download PDF](#) [View Record in Scopus](#) [Google Scholar](#)

[15] J. Holfeld, D. Zimpfer, K. Albrecht-Schgoer, A. Stojadinovic, P. Paulus, J.

Dumfarth, A. Thomas, D. Lobenwein, C. Tepekoylu, R. Rosenhek, W. Schaden, R. Kirchmair, S. Aharinejad, M. Grimm

**Epicardial shock-wave therapy improves ventricular function in a porcine model of ischaemic heart disease**

J. Tissue Eng. Regen. Med. (2014 May 19), [10.1002/term.1890](#)

[Epub ahead of print]

[Google Scholar](#)

[16] I. Gruenwald, B. Appel, N.D. Kitrey, Y. Vardi

**Shockwave treatment of erectile dysfunction**

Ther. Adv. Urol., 5 (2013), pp. 95-99

[CrossRef](#) [View Record in Scopus](#) [Google Scholar](#)

[17] C. Speed

**A systematic review of shockwave therapies in soft tissue conditions: focusing on the evidence**

Br. J. Sports Med., 48 (2014), pp. 1538-1542

[CrossRef](#) [View Record in Scopus](#) [Google Scholar](#)

[18] C.J. Wang

**An overview of shock wave therapy in musculoskeletal disorders**

Chang Gung Med. J., 26 (2003), pp. 220-232

[View Record in Scopus](#) [Google Scholar](#)






- [19] C.J. Wang, Y.C. Sun, T. Wong, S.L. Hsu, W.Y. Chou, H.W. Chang  
**Extracorporeal shockwave therapy shows time-dependent chondroprotective effects in osteoarthritis of the knee in rats**  
J. Surg. Res., 178 (2012), pp. 196-205  
[Article](#)  [Download PDF](#) [View Record in Scopus](#) [Google Scholar](#)
- [20] C.J. Wang, K.E. Huang, Y.C. Sun, Y.J. Yang, J.Y. Ko, L.H. Weng, F.S. Wang  
**VEGF modulates angiogenesis and osteogenesis in shockwave-promoted fracture healing in rabbits**  
J. Surg. Res., 171 (2011), pp. 114-119  
[Article](#)  [Download PDF](#) [View Record in Scopus](#) [Google Scholar](#)
- [21] C.J. Wang, Y.J. Yang, C.C. Huang  
**The effects of shockwave on systemic concentrations of nitric oxide level, angiogenesis and osteogenesis factors in hip necrosis**  
Rheumatol. Int., 31 (2011), pp. 871-877  
[CrossRef](#) [View Record in Scopus](#) [Google Scholar](#)
- [22] T.C. Yin, C.J. Wang, K.D. Yang, F.S. Wang, Y.C. Sun  
**Shockwaves enhance the osteogenetic gene expression in marrow stromal cells from hips with osteonecrosis**  
Chang Gung Med. J., 34 (2011), pp. 367-374  
[View Record in Scopus](#) [Google Scholar](#)
- [23] C.J. Wang, F.S. Wang, J.Y. Ko, H.Y. Huang, C.J. Chen, Y.C. Sun, Y.J. Yang  
**Extracorporeal shockwave therapy shows regeneration in hip necrosis**  
Rheumatology, 47 (2008), pp. 542-546  
[View Record in Scopus](#) [Google Scholar](#)
- [24] C.J. Wang, L.H. Weng, J.Y. Ko, J.W. Wang, J.M. Chen, Y.C. Sun, Y.J. Yang  
**Extracorporeal shockwave shows regression of osteoarthritis of the knee in rats**  
J. Surg. Res., 171 (2011), pp. 601-608  
[Article](#)  [Download PDF](#) [View Record in Scopus](#) [Google Scholar](#)
- [25] C.J. Wang, Y.C. Sun, K.K. Siu, C.T. Wu  
**Extracorporeal shockwave therapy shows site-specific effects in osteoarthritis of the knee in rats**  
J. Surg. Res., 183 (2013), pp. 612-619  
[Article](#)  [Download PDF](#) [View Record in Scopus](#) [Google Scholar](#)

- [26] C.J. Wang, C.Y. Huang, S.L. Hsu, J.H. Chen, J.H. Cheng  
**Extracorporeal shockwave therapy in osteoporotic osteoarthritis of the knee in rats: an experiment in animals**  
Arthritis Res. Ther., 16 (2014), p. R139  
[CrossRef](#) [Google Scholar](#)
- [27] Y.J. Chen, Y.R. Kuo, K.D. Yang, C.J. Wang, H.C. Huang, F.S. Wang  
**Shock wave application enhances pertussis toxin protein-sensitive bone formation of segmental femoral defect in rats**  
J. Bone Mineral Res.: Off. J. Am. Soc. Bone Mineral Res., 18 (2003), pp. 2169-2179  
[CrossRef](#) [View Record in Scopus](#) [Google Scholar](#)
- [28] F.S. Wang, K.D. Yang, Y.R. Kuo, C.J. Wang, S.M. Sheen-Chen, H.C. Huang, Y.J. Chen  
**Temporal and spatial expression of bone morphogenetic proteins in extracorporeal shock wave-promoted healing of segmental defect**  
Bone, 32 (2003), pp. 387-396  
[Article](#)  [Download PDF](#) [View Record in Scopus](#) [Google Scholar](#)
- [29] Y.J. Chen, Y.R. Kuo, K.D. Yang, C.J. Wang, S.M. Sheen Chen, H.C. Huang, Y.J. Yang, S. Yi-Chih, F.S. Wang  
**Activation of extracellular signal-regulated kinase (ERK) and p38 kinase in shock wave-promoted bone formation of segmental defect in rats**  
Bone, 34 (2004), pp. 466-477  
[Article](#)  [Download PDF](#) [View Record in Scopus](#) [Google Scholar](#)
- [30] S. Mayer-Wagner, J. Ernst, M. Maier, M. Chiquet, H. Joos, P.E. Muller, V. Jansson, B. Sievers, J. Hausdorf  
**The effect of high-energy extracorporeal shock waves on hyaline cartilage of adult rats in vivo**  
J. Orthop. Res.: Off. Publ. Orthop. Res. Soc., 28 (2010), pp. 1050-1056  
[View Record in Scopus](#) [Google Scholar](#)
- [31] D.M. Salter, W.H. Wallace, J.E. Robb, H. Caldwell, M.O. Wright  
**Human bone cell hyperpolarization response to cyclical mechanical strain is mediated by an interleukin-1beta autocrine/paracrine loop**  
J. Bone Mineral Res.: Off. J. Am. Soc. Bone Mineral Res., 15 (2000), pp. 1746-1755  
[CrossRef](#) [View Record in Scopus](#) [Google Scholar](#)
- [32] C. Eingartner, S. Coerper, J. Fritz, C. Gaissmaier, G. Koveker, K. Weise

**Growth factors in distraction osteogenesis. Immuno-histological pattern of TGF-beta1 and IGF-I in human callus induced by distraction osteogenesis**

Int. Orthop., 23 (1999), pp. 253-259

[View Record in Scopus](#) [Google Scholar](#)

- [33] D. Kaspar, C. Neidlinger-Wilke, O. Holbein, L. Claes, A. Ignatius  
**Mitogens are increased in the systemic circulation during bone callus healing**  
J. Orthop. Res.: Off. Publ. Orthop. Res. Soc., 21 (2003), pp. 320-325  
[Article](#)  [Download PDF](#) [View Record in Scopus](#) [Google Scholar](#)
- [34] T. Taniguchi, T. Matsumoto, H. Shindo  
**Changes of serum levels of osteocalcin, alkaline phosphatase, IGF-I and IGF-binding protein-3 during fracture healing**  
Injury, 34 (2003), pp. 477-479  
[Article](#)  [Download PDF](#) [View Record in Scopus](#) [Google Scholar](#)
- [35] P. Reher, M. Harris, M. Whiteman, H.K. Hai, S. Meghji  
**Ultrasound stimulates nitric oxide and prostaglandin E2 production by human osteoblasts**  
Bone, 31 (2002), pp. 236-241  
[Article](#)  [Download PDF](#) [View Record in Scopus](#) [Google Scholar](#)
- [36] F.S. Wang, C.J. Wang, S.M. Sheen-Chen, Y.R. Kuo, R.F. Chen, K.D. Yang  
**Superoxide mediates shock wave induction of ERK-dependent osteogenic transcription factor (CBFA1) and mesenchymal cell differentiation toward osteoprogenitors**  
J. Biol. Chem., 277 (2002), pp. 10931-10937  
[View Record in Scopus](#) [Google Scholar](#)
- [37] F.S. Wang, C.J. Wang, Y.J. Chen, P.R. Chang, Y.T. Huang, Y.C. Sun, H.C. Huang, Y.J. Yang, K.D. Yang  
**Ras induction of superoxide activates ERK-dependent angiogenic transcription factor HIF-1alpha and VEGF-A expression in shock wave-stimulated osteoblasts**  
J. Biol. Chem., 279 (2004), pp. 10331-10337  
[View Record in Scopus](#) [Google Scholar](#)
- [38] L. Martini, G. Giavaresi, M. Fini, P. Torricelli, M. de Pretto, W. Schaden, R. Giardino  
**Effect of extracorporeal shock wave therapy on osteoblastlike cells**  
Clin. Orthop. Relat. Res. (2003), pp. 269-280

[View Abstract](#)

Copyright © 2015 IJS Publishing Group Limited. Published by Elsevier Ltd. All rights reserved.

---



[About ScienceDirect](#)

[Remote access](#)

[Shopping cart](#)

[Advertise](#)

[Contact and support](#)

[Terms and conditions](#)

[Privacy policy](#)

We use cookies to help provide and enhance our service and tailor content and ads. By continuing you agree to the **use of cookies**.

Copyright © 2020 Elsevier B.V. or its licensors or contributors. ScienceDirect® is a registered trademark of Elsevier B.V.

ScienceDirect® is a registered trademark of Elsevier B.V.

